

Evaluation of several lolitrem-free endophyte/perennial ryegrass combinations

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Abstract

The fungal endophyte *Acremonium lolii*, in association with its ryegrass host, produces both peramine, a feeding deterrent to Argentine stem weevil, and lolitrem B, a neurotoxin causing ryegrass staggers. Endophyte strains vary in the ratio of production of these alkaloids in the ryegrass. In field evaluations of several endophyte/ryegrass combinations, 'Endosafe', a zero lolitrem B, high peramine strain protected its host against Argentine stem weevil attack and did not cause ryegrass staggers in lambs grazing three of four ryegrass cultivars. Lambs on the endophyte-free and Endosafe combinations had the highest liveweight gains and while endophyte-free ryegrass did not cause ryegrass staggers it suffered significant stem weevil damage. Ryegrasses with their 'wild-type' endophyte and a high lolitrem B strain 'Waiau', had only minor stem weevil damage but caused severe ryegrass staggers and weight loss in lambs. Ryegrass with the no lolitrem B, low peramine strain, '100A', suffered some stem weevil damage and although not causing ryegrass staggers, did cause weight loss in lambs. Ryegrass/endophyte metabolites in addition to lolitrem B may have affected animal performance on endophyte-infected ryegrass.

Keywords endophyte, Endosafe, perennial ryegrass, ryegrass staggers, Argentine stem weevil, liveweight gains, peramine, lolitrem B

Introduction

Resistance of perennial ryegrass (*Lolium perenne* L.) to Argentine stem weevil (*Listronotus bonariensis* Kuschel) attack and ryegrass staggers in grazing livestock are both associated with the endophyte *Acremonium lolii* (Latch, Christensen & Samuels) (Prestidge *et al.* 1982; Fletcher & Harvey 1981). The role of endophytes in pastoral agriculture in New Zealand has been reviewed recently by Fletcher *et al.* (1990).

The linking of one organism (in the same host) with two major problems in pastoral agriculture, and

the option of perennial ryegrass with or without endophyte, presented a dilemma for farmers in the drier regions. This option was available for any perennial ryegrass-based pasture, which comprise 75-80% of New Zealand's arable pastures. The industry in general either opted for cultivars with endophyte for better persistence, and accepted the associated animal health problems, or turned to alternative perennial grass species (Fletcher *et al.* 1990). Although ryegrass staggers and stem weevil resistance are both associated with the same host/endophyte relationship, they are mediated by different toxins (Rowan & Gaynor 1986; Gallagher *et al.* 1984). Peramine, the feeding deterrent to Argentine stem weevil, is a water-soluble pyrrolopyrazine alkaloid, which can be produced in culture (Rowan & Gaynor 1986). Lolitrem B, the main neurotoxin implicated in ryegrass staggers, is a lipid-soluble indole diterpene (Gallagher *et al.* 1984). Lolitrem B has so far been extracted only from ryegrass infected with *A. lolii*; it has not been produced by *A. lolii* in culture. This suggests the host ryegrass provides a precursor for, or at least plays a role in, metabolism of lolitrem B. Paxilline, a neurotoxin which produces symptoms similar to ryegrass staggers, is produced by *A. lolii* in culture and has also been proposed as a precursor to lolitrem B (Gallagher *et al.* 1977; Weedon & Mantle 1987).

Since the major endophyte effects (*viz.* ryegrass staggers and resistance to Argentine stem weevil attack) were mediated through two independent pathways the potential existed to exploit the benefits of endophyte by identifying or developing an endophyte which continued to produce peramine but produced zero or low levels of lolitrem B.

Such endophyte strains were isolated and introduced into several perennial ryegrasses using recently developed inoculation techniques (Latch & Christensen 1985).

The *A. lolii* ryegrass association produces another mammalian toxin, the ergopeptine alkaloid ergovaline (Bacon *et al.* 1986; Rowan & Shaw 1987; Rowan *et al.* 1990). This toxin has been implicated in the tall fescue toxicosis syndrome which occurs in cattle during summer in the US. The syndrome includes reduced liveweight gains and low serum prolactin levels. These

symptoms were also associated with the grazing of perennial **ryegrass** containing *A. lolii* (Fletcher & Barrell 1984). Since levels of ergovaline in **ryegrass** were similar to those recorded in the toxic tall fescue, the reduced prolactin levels, and possibly reduced liveweight gains found by Fletcher & Barrell (1984) were probably linked to ergovaline in infected ryegrass.

This paper reports on the results from the first field evaluation of several “new” **ryegrass** endophyte combinations, including those producing low levels of lolitrem B.

Materials and methods

A strain of endophyte, ‘Endosafe’ was isolated from **ryegrass** and grown in culture. Endosafe produces zero or very low levels of lolitrem B and ‘typical’ levels of **peramine** in its host. Two other endophyte strains, ‘100A’ and ‘Waiau’ (WA), were also isolated. 100A produces zero or low levels of lolitrem B and low levels of **peramine**. WA, from a North Canterbury **ryegrass** ecotype, produces very high levels of both lolitrem B and **peramine**.

Using the technique developed by Latch & Christensen (1985), Endosafe was inoculated into seedlings of four Grasslands perennial **ryegrass** cultivars: Pacific, Nui, Ruanui and Greenstone. WA was inoculated into all but Pacific; and 100A was inoculated into Greenstone and Ruanui only.

Seed from these plants was harvested in 1990 and sown into 126 m x 7 m plots of cultivated, ryegrass-free **Wakanui** silt loam at Lincoln. Endophyte-free seed of all four cultivars, and seed of Pacific and Nui with their ‘common’ or ‘wild type’ endophyte, were sown as controls. These 15 unreplicated combinations or treatments were sown with 200 kg/ha of DAP super-phosphate and without legume using a precision drill, at seeding rates of 3-10 kg/ha. Low seeding rates were necessitated by the limited availability of seed and to maximise the experimental area. A split application of urea (totalling 1.50 kg urea/ha) was made in spring.

The individually fenced plots were laxly grazed with **hoggets** for 14 days in spring to encourage the accumulation of basal plant tissue, increasing the potential for toxin production. With warm dry conditions the potential for **ryegrass** staggers and the expression of Argentine stem weevil damage was greatest for the summer/autumn grazing in February, when the trial was conducted.

Five-month-old Coopworth and Romney lambs with known staggers susceptibility were grazed on lucerne (non toxic) as one mob for 25 days until all were completely free of **ryegrass** staggers symptoms. Having had recent serious **ryegrass** staggers, the time required for toxins to reach threshold levels in the lambs was

reduced and the potential for **ryegrass** staggers to occur on the trial (Fletcher 1983) was maximised.

The lambs were fasted for 24 hours, weighed, then allocated to treatments so that all 15 mobs had a similar mean weight per head, each mob had a similar quantity of **herbage** on offer per head and each mob had a similar balance of the two breeds. Lambs averaged 9 per treatment.

Conditions during the 6 weeks before grazing were warm and dry. However, at the start of grazing 50 mm of rain fell in 2 days, resulting in **ryegrass** growth rates of 50-60 kg/DM/day. To cope with this increased growth and to reduce the time for grazing heights to reach the lower ‘toxic zone’, ewes were **introduced** (in proportion to lamb numbers in each treatment) for 5 days. On all treatments grazing height was 30 mm or below for the last 7 days’ grazing, maximising intake of toxic material.

Lambs were weighed 14 days after grazing began and again after 28 days when the trial concluded. Each weighing was preceded by a 24-hour fast. Lambs were scored individually for incidence and severity of **ryegrass** staggers on 5 occasions during the trial using a modified Keogh scale (Keogh 1973).

The quantity of **herbage** present on each treatment before grazing and on 5 occasions during grazing was measured with a capacitance probe, calibrated against **quadrats** cut to ground level. Randomly selected **herbage** samples, cut at ground level, were taken from each treatment at the start, middle and at end of the trial. These were then freeze dried and analysed for lolitrem B, ergovaline and **peramine** content (Rowan & Shaw 1987; Tapperet *al.* 1989). Fresh samples were taken at the start of the trial; stained epidermal strips from leaf sheaths were examined under a light microscope (Latch & Christensen 1985) to determine the percentage of tillers with endophyte.

Argentine stem weevil numbers (adults and larvae) were assessed for each treatment in September and December. Larval numbers were assessed from randomly selected tillers in December using the method of Goldson (1978). Adult and larval damage was scored in March on 100 tillers randomly selected from each treatment. Larval damage were recorded as light, moderate or severe.

Although the trial was neither balanced nor fully replicated, the effect of endophyte strain and **ryegrass** cultivar were tested statistically using the interaction as an error term.

Results

In Greenstone WA and 100A treatments, 60 and 80% respectively of tillers were infected with endophyte. Apart from endophyte-free treatments, which had no

endophyte. all other treatments had more than 95% of tillers infected with endophyte.

Endophyte strain had a significant effect on **ryegrass** staggers ($P<0.01$) at all observations (Figure 1). Lambs on WA and wild-type treatments were more affected than those on **100A**, Endosafe and endophyte-free. Lambs on the **Ruanui/Endosafe** treatment showed mild **ryegrass** staggers despite the absence of lolitrem B.

Endophyte strain significantly affected liveweight gain over the total trial period ($P<0.01$) (Figure 2), where on average lambs on endophyte-free and Endosafe treatments gained weight **while** those on other treatments lost weight. Lambs on **100A**, Endosafe and common endophyte treatments did not differ significantly for the first 14 days. These treatments gained less weight than endophyte-free treatments but more than for WA.

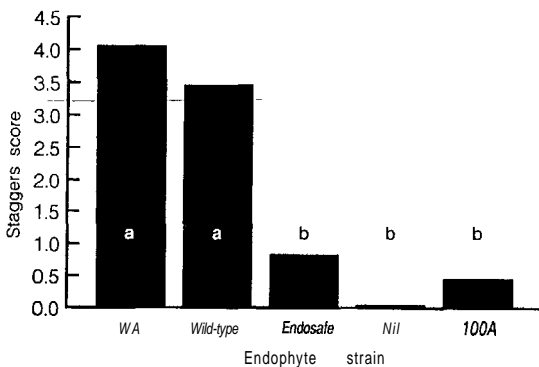


Figure 1 Ryegrass staggers scores during February-March for lambs grazing perennial ryegrass with different endophyte treatments. (Means of 5 scores over 28 days). Treatments with the same letters are not significantly different.

During the second 14 days **herbage** availability limited liveweight gain to various degrees, particularly on some endophyte-free treatments, as a result of the higher palatability of these treatments and consequent higher intake during the first period. All animals lost weight during the second 14-day period as **herbage** availability decreased.

The presence of endophyte significantly reduced Argentine stem weevil damage. The percentage of tillers with adult feeding was significantly higher in the nil endophyte treatment than in the endophyte-infected treatments in March (Figure 3.). Number of tillers damaged by adult feeding did not differ significantly in the wild-type and **100A** treatments. Larval densities recorded in December were approximately 1.8 times higher on the nil endophyte treatments than on Endosafe, WA or wild-type treatments, with numbers on **100A** intermediate between these groups. Tiller damage by larvae followed a similar pattern, with a mean of 8.5% of tillers from

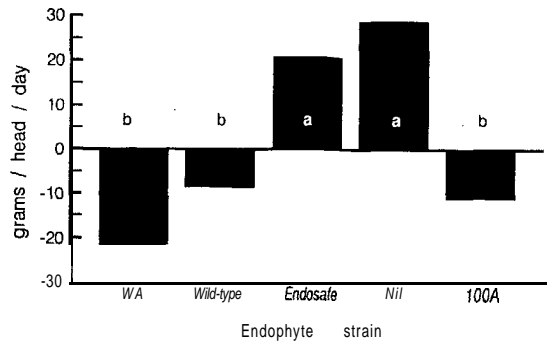


Figure 2 Mean liveweight change in lambs grazing perennial ryegrass with different endophyte treatments. Treatments with the same letters are not significantly different.

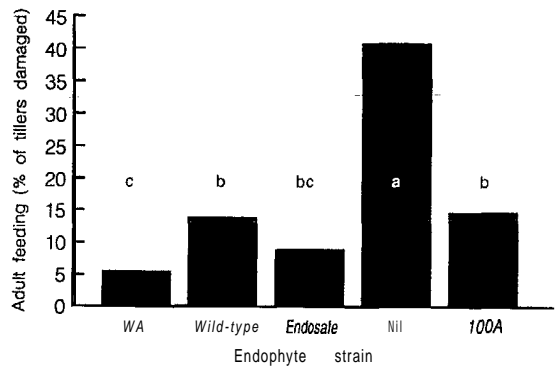


Figure 3 Perennial ryegrass tillers (as % of total tillers) damaged by Argentine stem weevil adults on perennial ryegrass with different endophyte treatments. Treatments with the same letters are not significantly different.

the endophyte-free plots damaged compared **with 3.6%** from the infected plots. Tiller damage was much less severe when endophyte was present. A cultivar effect was also apparent, with Greenstone showing more adult and larval attack than other cultivars.

Mean **peramine** and lolitrem B concentrations, in field samples collected on 5 occasions throughout the trial (Table 1) were similar to those in the original ryegrass plants **innoculated** with endophyte. They were also consistent with the patterns of stem weevil damage and **ryegrass** staggers (Figures 1 and 3).

Table 1 Effect of endophyte strain on peramine and lolitrem B concentrations (ppm), means from five sample dates.

	Peramine	Lolitrem B
W A	17.7	8.0
Wild-type	10.5	4.7
Endosafe	11.3	0.1
100A	4.2	0
Endophyte-free	0.1	0

Discussion

All cultivars with Endosafe showed good resistance to Argentine stem weevil compared with wild-type and endophyte-free combinations, and three of the four Endosafe combinations did not cause any significant ryegrass staggers in grazing lambs in summer. This is a significant finding for the pastoral industry, but mild staggers in lambs grazing Ruanui with Endosafe suggests caution may be needed in extrapolating these results to other ryegrass cultivars inoculated with Endosafe. Lambs grazing Ruanui with WA had more severe staggers than other cultivars with the same endophyte. Significant effects of host cultivar on the development of ryegrass staggers have recently been demonstrated (Fletcher & Sutherland unpubl.). As lolitrem B cannot be produced from 'normal' endophyte culture, only by the endophyte in conjunction with its host, the host ryegrass may produce a precursor to lolitrem B. These indications of possible interaction between cultivar, or host plant genotype, and endophyte must be further investigated.

Regression of lolitrem B concentrations against ryegrass staggers using data from all treatments established that lolitrem B concentration is closely correlated with ryegrass staggers ($r = 0.92$) However, Ruanui with Endosafe had zero lolitrem B at all samplings yet caused mild, but significant, ryegrass staggers. This suggests other tremorgens may be involved, e.g. paxilline (Gallagher *et al.* 1977; Weedon & Mantle 1987; Gallagher & Prestidge 1990).

Our results agree with previously published reports on the effect of wild-type endophyte and endophyte-free ryegrass on liveweight (Fletcher *et al.* 1990). The weight loss in lambs on the lolitrem B-free 100A treatments suggests toxins, present in endophyte-infected ryegrass, in addition to lolitrem B, may influence liveweight change, e.g. ergopeptine alkaloids (Fletcher & Barrell 1984; Bacon *et al.* 1986; Rowan *et al.* 1990; Piper & Fletcher 1990).

The Waiiau, common and Endosafe endophytes all conferred a high degree of resistance to stem weevil. Plants with the 100A strain also showed good resistance despite lower concentrations of the feeding deterrent peramine (Table 1). Levels of larval damage in March were relatively low, but damage would probably have been considerably higher during summer.

Although Endosafe has successfully demonstrated that the benefits of endophytes can be exploited while overcoming animal health problems, future research needs to evaluate the role of known, and any as yet unidentified, plant/endophyte metabolites and examine the role of the host plant in toxin production and manifestation of effects on animal health and performance.

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REFERENCES

- Bacon, C.W.; Lyons, P.C.; Porter, J.K.; Robbins, J.D. 1986. Ergot toxicity from endophyte-infected grasses: A review. *Agronomy journal* 78: 106-116.
- Fletcher, L.R. 1983. Effects of presence of *Lolium* endophyte on growth rates of weaned lambs, growing on to hoggets, on various ryegrasses. *Proceedings of the NZ Grassland Association* 44: 237-239.
- Fletcher, L.R.; Barrell, G.K. 1984. Reduced liveweight gains and serum prolactin levels in hoggets grazing ryegrasses containing *Lolium* endophyte. *NZ Veterinary Journal* 32: 139-140.
- Fletcher, L.R.; Harvey, I.C. 1981. An association of a *Lolium* endophyte with ryegrass staggers. *NZ Veterinary Journal* 29: 185-186.
- Fletcher L.R.; Hoglund J.H.; Sutherland B.L. 1990. The impact of Acremonium endophytes in New Zealand, past, present and future. *Proceedings of the NZ Grassland Association* 52: 227-235.
- Gallagher, R.T.; Hawkes, A.G.; Steyn, P.S.; Vleggaar, R. 1984. Tremorgenic neurotoxins from perennial ryegrass causing ryegrass staggers disorder of livestock: structure and elucidation of lolitrem B. *Journal of the Chemical Society, chemical communications (London)*: 614-616.
- Gallagher, R.T.; Keogh, R.G.; Latch, G.M.C.; Reid, C.S.M. 1977. The role of fungal tremorgens in ryegrass staggers. *New Zealand Journal of Agricultural research* 20: 431-430.
- Gallagher, R.T.; Prestidge, R.A. 1990. Structure-activity studies on indole diterpenes, including lolitrems and related indoles and tremorgens. *Proceedings of the 1st International Symposium on Acremonium/Grass Interactions*. pp 80-81.
- Goldson S.L. 1978. Simple technique for extracting Argentine stem weevil (*Hyperodes bonariensis*) larvae from ryegrass tillers (*Coleoptera: Curculionidae*). *The New Zealand Entomologist* 6:437. Keogh, R.G. 1973. Induction and prevention

-
- of ryegrass staggers in grazing sheep. *NZ Journal of Experimental Agriculture 1*: 55-57.
- Latch, G.C.M.; Christensen, M.J. 1985. Artificial infection of grasses with endophytes. *Annals of Applied Biology 107*: 17-24.
- Piper, E.L.; Fletcher, L.R. 1990. Influence of adopamine antagonist on ryegrass staggers. *Proceedings of the 1 st International Symposium on Acremonium/Grass Interactions*. pp 248-249.
- Prestidge, R.A.; Pottinger, R.P.; Barker, G.M. 1982. An association of *Lolium* endophyte with ryegrass resistance to Argentine stem weevil. *Proceedings of the 35th NZ Weed and Pest Control Conference*: 119-122
- Rowan, D.D.; Gaynor, D.L. 1986. Isolation of feeding deterrents against Argentine stem weevil from ryegrass infected with *Acremonium lolii*. *Journal of Chemical Ecology 12*: 647-658.
- Rowan, D.D.; Shaw, G.J. 1987. Detection of ergopeptide alkaloids in endophyte-infected perennial ryegrass by tandem mass spectrometry. *NZ Veterinary Journal 3.5*: 197-198.
- Rowan D.D.; Tapper B.A.; Sergejew N.L.; Latch G.C.M. 1990. Ergopeptide alkaloids in endophyte-infected ryegrasses and fescues in New Zealand. *Proceedings of the 1 st International Symposium on Acremonium/Grass Interactions*. pp 97-99.
- Tapper B.A.; Rowan D.D.; Latch G.C.M. 1989. Detection and measurement of the alkaloid peramine in endophyte-infected grasses. *Journal of Chromatography 463*: 133-138.
- Weedon, C.M.; Mantle, P.G. 1987. Paxilline biosynthesis by *Acremonium loliae*; a step towards defining the origin of lolitrem neurotoxins. *Phytochemistry 26*: 969-971.